## METHODS OF DETECTION OF SPECIFIC ANTIBODIES IN BRUCELLOSIS

*Djabbarova N.R.-* assistant of the department of clinical laboratory diagnosis with the course of clinical laboratory diagnostics of PGD;

**Daminov F.A.** – DSc, head of the department of clinical laboratory diagnosis with the course of clinical laboratory diagnostics of PGD;

**Xolmatova D.** - cadet of the department of clinical laboratory diagnosis with the course of clinical laboratory diagnostics of PGD;

**Rustamova F.-** cadet of the department of clinical laboratory diagnosis with the course of clinical laboratory diagnostics of PGD; Samarkand state medical university Samarkand, Uzbekistan

**Plate Agglutination Reaction (Heddleston Reaction).** The advantage of this method is the simplicity of setting up the reaction, quick results, and the sensitivity of the reaction. A single brucellosis diagnosticum is used as an antigen to perform the Heddleston and Wright reactions. In the case of a positive reaction, flakes (agglutinate) appear in the drops of serum containing the antigen in the very first minutes. The maximum observation period is 8 minutes. For the diagnosis of brucellosis, only a positive result of the reaction is important. In cases of uncertain or negative results, along with epidemiological or epizootic indications, or in hospitals and donor screenings where it is necessary to determine the titer of agglutinins and their dynamics, the Wright, Coombs, Passive Hemagglutination Reaction (PHAR), and Enzyme-Linked Immunosorbent Assay (ELISA) should be used [1,4,6,8,9].

**Tube Agglutination Reaction (Wright Reaction).** The agglutination reaction is one of the main methods for diagnosing brucellosis in humans. It holds the most diagnostic value in acute and subacute forms of brucellosis. A diagnostic titer is considered to be an agglutination reaction of at least 2+ at a serum dilution of 1:100 or higher [7,8,9].

Antiglobulin Test (Coombs Reaction). In diagnosing brucellosis in humans and animals, especially with chronic infections where the agglutination reaction may be negative or positive at low titers, it is important to identify incomplete antibodies. The Coombs reaction uses a pre-titrated antiglobulin serum against human globulins (for example, antiglobulin serum for immunoelectrophoresis against human globulins). A diagnostic titer in the Coombs test is considered to be agglutination of at least 2+ at a serum dilution of 1:50 [1,2,3].

**Passive Hemagglutination Reaction (PHAR).** PHAR is a specific and highly sensitive method for detecting brucellosis antibodies in human serum. The serum titer is determined by the last dilution that yields a reaction of at least 3+. Interpretation of



PHAR results in humans: a titer of 1:50 is considered doubtful, while a titer of 1:100 and higher is considered positive [10,11,12].

**Enzyme-Linked Immunosorbent Assay (ELISA).** This method is used to diagnose all forms of the disease and during epidemiological screening of the population, as well as when selecting individuals for vaccination against brucellosis. A diagnostic ELISA test system is used to determine brucellosis antibodies. Specific antibodies in human serum are detected through the interaction of brucellosis antigen (LPS), adsorbed on a polystyrene flat-bottom plate, with the antibodies of the tested serum. A diagnostic titer in ELISA is considered a serum dilution of more than 1:400 [13,14,15].

## Tests Revealing Increased Sensitization to Brucellosis Antigen.

Burnet Skin Allergy Test (Intradermal Allergy Test): This test is based on the ability of the body sensitized to brucellosis antigen to respond specifically with a local reaction (swelling, pain) to the intradermal administration of brucellosis allergen. The reaction is specific but appears in patients later than antibodies and persists for a long time, sometimes for years, after the clinical symptoms disappear. It should be noted that a positive allergic reaction may occur in cases of asymptomatic infection, as well as in individuals vaccinated with live brucellosis vaccine and those who have had long-term contact with the specific antigen [16,17,18].

**Leukocyte Lysis Reaction.** The introduction of a specific antigen into a sensitized organism is significant for the examined individual. Therefore, an effective method for detecting delayed hypersensitivity in vitro is the leukocyte lysis reaction (LLR). LLR is based on the registration of the destruction of leukocytes in a sensitized organism under the influence of a specific antigen, recorded in vitro. LLR has strict specificity, allows quantitative assessment of the degree of sensitization, and provides a result 3-4 hours after blood sampling [19,20,21].

**Procedure for Leukocyte Lysis Reaction (LLR).** LLR is performed in test tubes made of chemically pure glass. The antigen used is a suspension of heat-killed brucella (a vaccine strain B. abortus 19BA may be used) at a concentration of  $1 \times 10^{7}$  µl/ml. Blood for testing is collected in a volume of 1 ml and placed in a tube with heparin at a rate of 75-80 IU of heparin per 1 ml of blood. The specific leukocyte lysis index (SLLI) is calculated by determining the difference - the percentage of leukocyte reduction in the experimental tube minus the percentage of leukocyte reduction in the control. SLLI is expressed as a negative value and ranges from -10% to -30%. An SLLI less than -10% indicates nonspecific lysis [1,2,3].

It is advisable to highlight the laboratory test complexes used for diagnosing brucellosis based on the goals and level of medical care organization:

1. For epidemiological screening of populations in outbreak areas, the following are used: agglutination reaction (Heddleston reaction), Wright reaction, PHAR, ELISA,

and the Burnet skin allergy test.

2. For population screening before preventive vaccination: the Heddleston reaction or ELISA, the Burnet skin allergy test, or the leukocyte lysis reaction.

3. For diagnosing acute and subacute brucellosis: bacteriological studies, agglutination reaction, PHAR, and ELISA are performed. In cases of negative results, the Coombs reaction is used.

4. For diagnosing chronic brucellosis and during dispensary observation of individuals who have recovered from brucellosis: the Coombs reaction, ELISA, PHAR, and skin allergy tests are recommended.

Serological reactions and the skin allergy test have different diagnostic significance at various stages of the disease and cannot replace each other. This necessitates the use of a comprehensive sero-allergic method, which is the most reliable way to diagnose brucellosis. In the early stages of the disease (within the first 6 months), the diagnostic value of serological methods is higher than that of allergic methods; serological reactions during this period are positive in almost 98% of cases. As the duration of the disease increases, the percentage of positive serological reactions (agglutination reaction, PHAR) begins to decline. In the later stages of the disease, the Coombs reaction, ELISA, and the intradermal allergy test have greater diagnostic value [4,5,6].

It is important to consider that while high antibody titers almost always indicate the presence of infection, low antibody titers or their complete absence do not exclude the possibility of disease. Therefore, repeated testing at intervals of 1-2 weeks is recommended, especially when acute brucellosis is suspected. It should be noted that a positive agglutination reaction with brucellosis antigen may also be given by sera containing antibodies to microorganisms that have common antigenic determinants with brucella [3,4,5,6].

## **References:**

1. Kudratova Z. E. et al. Current modern etiology of anemia //Open Access Repository.  $-2023. - T. 10. - N_{2}. 10. - C. 1-4.$ 

2. Burxanova D. S., Umarova T. A., Kudratova Z. E. Acute myocarditis linked to the administration of the COVID 19 vaccine //Центральноазиатский журнал образования и инноваций. – 2023. – Т. 2. – №. 11. – С. 23-26.

3. Кудратова 3. Э. и др. Атипик микрофлора этиологияли ўткир обструктив бронхитларининг ў зига хос клиник кечиши //Research Focus. - 2022. - Т. 1. - №. 4. - С. 23-32.

4. Kudratova Z. E, Normurodov S. Etiological structure of acute obstructive bronchitis in children at the present stage - Thematics Journal of Microbiology, 2023. P.3-12.

 Kudratova Z. E., Tuychiyeva S. K. Atipik mikroflora etiologiyali o'tkir obstruktiv bronxitlar etiopatogenezining zamonaviy jixatlari. Research Focus, 2023, B. 589-593.
Kudratova Z. E., Karimova L. A. Age-related features of the respiratory system. Research Focus, Tom 2, P. 586-588.

7. Исомадинова Л. К., Даминов Ф. А. Современная лабораторная диагностика



хронического пиелонефрита у детей //Journal of new century innovations. – 2024. – Т. 49. – №. 2. – С. 112-116.

8. Isomadinova L. K., Daminov F. A. Glomerulonefrit kasalligida sitokinlar ahamiyati //Journal of new century innovations. – 2024. – T. 49. – №. 2. – C. 117-120.

9. Isomadinova L. K., Qudratova Z. E., Shamsiddinova D. K. Samarqand viloyatida urotiliaz kasalligi klinik-kechishining o'ziga xos xususiyatlari //Центральноазиатский журнал образования и инноваций. – 2023. – Т. 2. – №. 10. – С. 51-53.

10. Isomadinova L. K., Qudratova Z. E., Sh B. F. Virusli gepatit b fonida Covid-19 ning klinik laborator kechish xususiyatlari //Journal of new century innovations. –  $2023. - T. 30. - N_{\odot}. 3. - C. 60-65.$ 

11. Isomadinova L. K., Yulayeva I. A. Buyraklar kasalliklarning zamonaviy diagnostikasi //Центральноазиатский журнал образования и инноваций. – 2023. – Т. 2. – №. 10 Part 3. – С. 36-39

12. Kudratova Zebo Erkinovna, Tamila Abdufattoevna Umarova, & Sirojeddiova Sanobar. (2024). Modern types of immunoenzyme analysis methods old problems. Web of Discoveries: Journal of Analysis and Inventions, 2(6), 67–70.

13. Набиева Ф. С., Мусаева Ф.Р. Лабораторная диагностика острого гломерулонефрита //Journal of new century innovations. – 2023. – Т. 30. – №. 3. – С. 150-152.

14. Жаббарова Д.З., Набиева Ф.С., Якубова Д. М. Применение иммуноферментного анализа в медицине //TADQIQOTLAR. – 2024. – Т. 46. – №. 1. – С. 40-42.

15. Чориева Т.А., Якубова Д.М., Набиева Ф.С. Диагностика и профилактика torch инфекции у беременных //TADQIQOTLAR. – 2024. – Т. 46. – №. 1. – С. 26-30.

16. Mamatova M. N. Study of the biological properties of rabies by the method of diagnosis of the "gold standard" //GOLDEN BRAIN.  $-2024. - T. 2. - N_{2}. 4. - C. 129-144.$ 

17.Юлаева И. А., Расулова М. Р., Шодиев Ж. Х. Переломы костей носа в практике судебномедицинской экспертизы // Eurasian journal of medical and natural sciences.- 2023. - Т. 1. - № 1. - С. 78-84.

18.Юлаева И. А., Расулова М. Р., Шодиев Ж. Х. Современная классификация переломов костей носа // "Talqin va tadqiqotlar" ilmiy-uslubiy jurnali. – 2023. - № 17. – С. 120-127.

19.Юлаева И. А., Расулова М. Р., Шодиев Ж. Х. Современные методы диагностики переломов костей носа // "Talqin va tadqiqotlar" ilmiy-uslubiy jurnali. – 2023. - № 17. – С. 225-235.

20. Yulaeva I. A., Ibragimova N. S. Combination of transitor hyperammoniemia with post-hypoxic syndrome in newborn // "World Bulletin of Public Health (WBPH)" Scholar Express Journals– 2023.- T. 21. – C. 12-14.

21. Юлаева И.А., Ибрагимова Н.С. Патогенетические, клинико-лабораторные и инструментальные аспекты диагностики синдрома поликистозных яичников у женщин репродуктивного возраста // JOURNAL OF NEW CENTURY INNOVATIONS – 2023. – Т. 26. - №3. – С. 180-184.